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## Improving Results of Flexor Tendon Repair and Rehabilitation

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**Background:** The global time and effort attributed to improving outcomes in the management of flexor tendon injury are large, but the degree of advancement made over the past 50 years is relatively small. This review examines the current perceived wisdom in this field and aims to explore the limitations to the authors' understanding of the tendon healing process, examining how this may be a factor that has contributed to the authors' modest progress in the field.

**Methods:** The authors critically evaluate the sum of laboratory and clinical literature on the topic of zone II flexor tendon management that has guided their practice and provide evidence to support their methods.

**Results:** The review highlights some of the key developments over the years and assesses their influence on changing current practice. It also highlights recent innovations, which have the potential to influence flexor tendon outcomes by altering the surgical approach, techniques, and rehabilitation regimens. Future innovations in the field will also be discussed to examine their potential in expanding the development in the management of flexor tendon injury.

**Conclusions:** A better understanding of flexor tendon biology will allow progress in developing new therapies for flexor tendon injuries; however, there are as yet few real breakthroughs that will dramatically change current practice. (*Plast. Reconstr. Surg.* 134: 913e, 2014.)

The management of flexor tendon injuries remains one of the most published topics in hand surgery, with the numbers of publications on this subject seeing a year-on-year increase (Fig. 1). The perfect repair and outcome continue to evade us,<sup>1</sup> despite the flexor tendon repair being one of the earliest skills acquired as a hand surgeon in either plastic or orthopedic surgery training.<sup>2</sup> New tendon repairs and hand therapy regimens are reported regularly. Occasionally, there is an announcement of a new treatment modality that promises hope for this clinical conundrum, but this rarely becomes part of standard practice. Over the past 50 years, there have been many innovations, but overall outcomes have not changed dramatically. For example, the best series published in the 1970s showed that a two-strand repair with

simple circumferential suture and a Kleinert type rehabilitation regimen had a 5 percent rupture rate, with 75 percent of patients achieving good to excellent functional outcomes in 28 zone II injuries.<sup>3</sup> This compares favorably with more recent studies showing that a four-strand repair and early active mobilization regimen had a 5 percent rupture rate, with 71 percent achieving good to excellent outcomes in 73 cases.<sup>4</sup>

Real paradigm shifts in this area require us to rethink the whole process of flexor tendon biology

**Disclosure:** *Neither author has a financial interest in any of the products or devices mentioned in this article.*

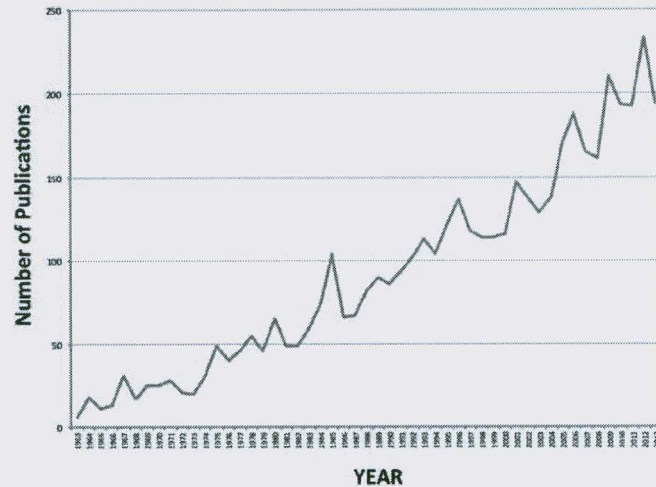
Supplemental digital content is available for this article. Direct URL citations appear in the text; simply type the URL address into any Web browser to access this content. Clickable links to the material are provided in the HTML text of this article on the *Journal's* Web site ([www.PRSJournal.com](http://www.PRSJournal.com)).

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**Fig. 1.** PubMed statistics on flexor tendon publications over the past 50 years.

and how this relates to the clinical picture. This review aims to present the current best evidence of how to manage flexor tendon injury, but also challenges these surgical and rehabilitation management strategies with alternative approaches that may potentially see further improvement.

### SCALE OF THE PROBLEM

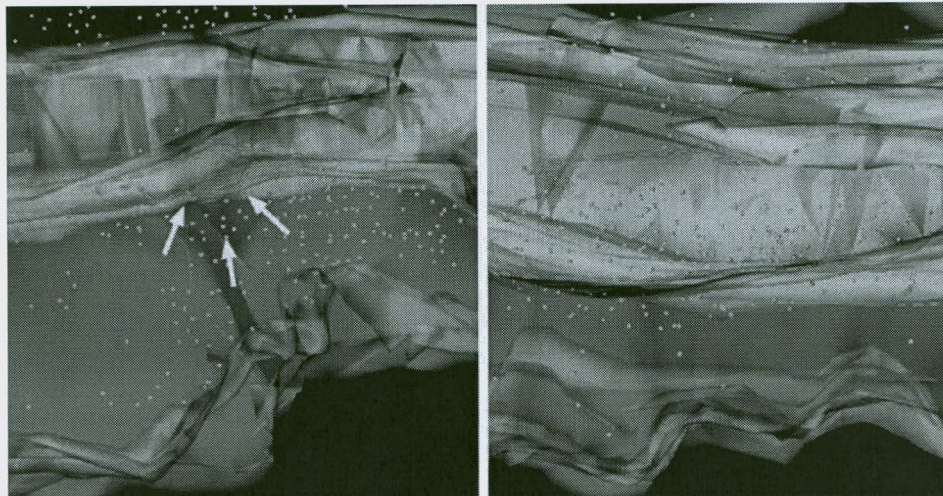
Flexor tendon injuries are increasing in frequency on an annual basis in the United Kingdom, with there being an estimated 18,000 primary repairs performed per year, equating to an incidence of 41 per 100,000. Approximately 2000 per year require secondary tenolysis or two-stage flexor tendon grafting, suggesting an 11 percent reoperation rate.<sup>5</sup> Through meta-analysis of systematic reviews,<sup>6</sup> complications have been reported as 4 percent rupture rates, 6 percent reoperation rates, and 4 percent adhesion rates, which suggest that the problem is underreported. Of these secondary operations, 58 percent had tenolysis, 38 percent had simple rerepair, and 4 percent had tenolysis and rerepair. Ninety-one percent had one reoperation, whereas 8 percent had two further operations and 1 percent had three.

### FLEXOR TENDON HEALING PARADIGM

Despite better scientific understanding of how tendon heals, the clinical community still relies on concepts that were introduced 50 years ago. The concepts of intrinsic and extrinsic healing were largely borne of observations relating to the experimental design and the modality being used

to investigate the biology at the time. For example, Potenza's original experiments<sup>7</sup> involved using a tube to interrupt adhesion growth into the tendon in canine forepaws, which led to tendon necrosis caused by lack of an extrinsic blood supply. This led to the notion of an "extrinsic" healing process, whereby adhesions were necessary to revascularize the tendon. A critical explanation would suggest that insertion of silicone foreign material led to a significant inflammatory response, increasing sheath and tissue pressure within the digit and thereby contributing to tendon necrosis. Studies over the years have shown that using other barrier methods to inhibit adhesions actually has little bearing on how the tendon heals, other than to reduce adhesions.<sup>8</sup> Matthews and Richards<sup>9</sup> found that retracting the tendon out with the sheath and wounding the tendon led to healing in the absence of adhesion formation, indicating that repair could be initiated by cell populations "intrinsic" to the tendon. Studies have supported these findings by showing injured intrasynovial tendon healed independently of adhesions in a synovial environment,<sup>10</sup> in culture<sup>11,12</sup> and when transplanted into subcutaneous diffusion chambers.<sup>13</sup> Cells provided with the appropriate nutrition in culture will behave in this fashion, which is not unique phenomenology to tendon. Like most cells, tendon fibroblasts will proliferate and spread, smoothing over any rough surface of tissue in the process although, uniquely, when given two points of tension, tendon fibroblasts will self-assemble into three-dimensional structures.<sup>14,15</sup>

The reality of tendon healing is far more complex, involving a careful interplay of cells, matrix, gene expression, and growth factors between the



**Fig. 2.** Three-dimensional cell mapping of adhesion formation in zone II of the mouse flexor tendon. (Left) At 7 days after injury, early formation of tendon adhesion (outlined in dotted red) at the interface between tendon and surrounding tissue. Collagen synthesizing cells are shown in green. (Right) At 21 days after injury, there is established adhesion formation with lots of collagen being deposited at the interface between tendon and surrounding tissue.

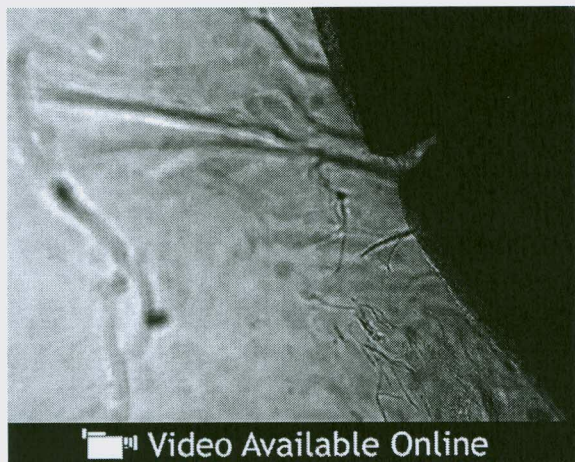
tendon and the surrounding structures.<sup>16,17</sup> The tendon healing paradigm is similar to that of wound healing but must be taken in context with the dynamic gliding anatomy of the tissues and digits.<sup>18</sup>

Immobilizing flexor tendon injuries allows for healing to occur between the two injured tissues, which overlap during the healing process. Our observations on a simple murine flexor tendon injury model<sup>16</sup> showed some resemblance to early experiments by Lindsay and McDougall,<sup>19</sup> who found that the cells from both subcutaneous tissues, sheath and tendon, were active in cell migration, collagen synthesis, and cellular proliferation. It was also apparent that the inherent swelling caused by injury allows the tendon and sheath to come into contact while the healing process begins (Fig. 2). [See Video, Supplemental Digital Content 1, which displays a zone II flexor tendon adhesion formation in a mouse (using three-dimensional cell mapping). (A - Partial Laceration) After the initial partial laceration of tendon and tissues. (B - Inflammatory Phase) The inflammatory phase is initiated and a big influx of inflammatory cells occur (red). The tissues begin to swell, which brings the tendon and surrounding tissues into contact and the initiation of collagen synthesis (green) begins in the subcutaneous tissues. Macrophages are recruited into the adhesion (yellow). (C - Early Proliferative Phase) Gradually, the adhesion-forming process increases with maximum synthetic activity and cellular infiltration at approximately 3 weeks.

(D - Established Synthetic Phase) Gradually the tissue swelling and cell number decrease through apoptosis and the adhesion remodels leaving a



**Video 1.** Supplemental Digital Content 1 displays a zone II flexor tendon adhesion formation in a mouse (using three-dimensional cell mapping). (A - Partial Laceration) After the initial partial laceration of tendon and tissues. (B - Inflammatory Phase) The inflammatory phase is initiated and a big influx of inflammatory cells occur (red). The tissues begin to swell, which brings the tendon and surrounding tissues into contact and the initiation of collagen synthesis (green) begins in the subcutaneous tissues. Macrophages are recruited into the adhesion (yellow). (C - Early Proliferative Phase) Gradually, the adhesion-forming process increases with maximum synthetic activity and cellular infiltration at approximately 3 weeks. (D - Established Synthetic Phase) Gradually the tissue swelling and cell number decrease through apoptosis and the adhesion remodels leaving a thinner adhesion (E - Remodelling and Resolution Phase). This video is available in the "Related Videos" section of the full-text article on PRSJournals.com or available at <http://links.lww.com/PRS/B157>.



**Video 2.** Supplemental Digital Content 2 displays a partial tendon laceration in vitro. A mouse flexor tendon is partially divided and time lapsed over the course of 24 hours, <http://links.lww.com/PRS/B158>.

thinner adhesion (*E- Remodelling and Resolution Phase*). This video is available in the “Related Videos” section of the full-text article on PRSJournals.com or available at <http://links.lww.com/PRS/B157>.] The tendon and sheath are covered by an epithelium that acts as a barrier to the migration of cells which, when breached, allows for cells to move freely along a growth factor gradient.<sup>20</sup> (See Video, Supplemental Digital Content 2, which displays a partial tendon laceration in vitro. A mouse flexor tendon is partially divided and time lapsed over the course of 24 hours, <http://links.lww.com/PRS/B158>.)

Mechanism of injury is important in predicting the degree of damage signaling in the tissues. For example, a sharp clean cut will heal well if the repair is performed carefully; however, the trauma from a tearing, crush, avulsion, aggressive coagulation, infection, or overzealous handling will lead to a greater injury response, which in turn dictates adhesion formation (Table 1).<sup>21–24</sup> In the clinical setting where the zone II flexor

tendon had tearing and saw injuries, significantly poorer functional outcomes were observed compared with those with sharp clean injuries to their flexor tendons (range of movement,  $86 \pm 14$  degrees versus  $114 \pm 7$  degrees, respectively;  $p = 0.05$ ).<sup>25</sup> In vivo studies in canines have also shown that suture techniques that exhibit more suture on the tendon surface, such as the Massachusetts General Hospital Repair, generate more friction, presumably through adhesions, than suture techniques such as the modified Kessler repair that exhibit less suture on the tendon surface<sup>26</sup>; thus, a balance must be made between gaining adequate strength without overtraumatizing the tendon<sup>27</sup> or producing too much glide resistance.<sup>28</sup>

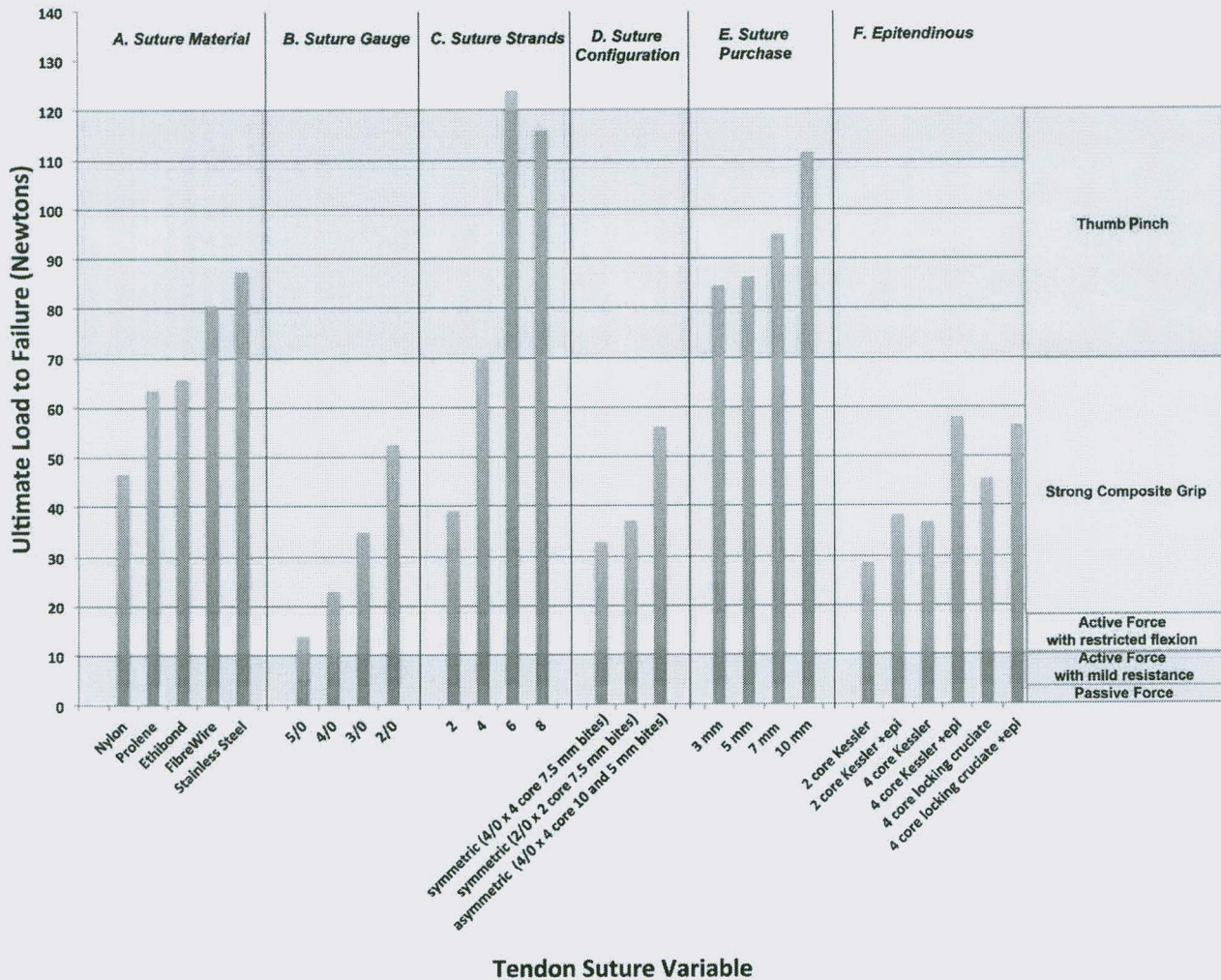
## TENDON REPAIR

Hand therapy, with early active mobilization, has proved to be the main effective modality in preventing adhesions<sup>29</sup>; thus, surgically, the aim has been to make our tendon repairs strong. Any repair should take into account the forces acting on the finger during rehabilitation.<sup>30</sup> Strength can be gained by suture material, gauge of suture, number of core strands, suture purchase, suture configuration, and addition of an epitendinous suture (Fig. 3).

The ideal suture is deemed to be strong, minimally reactive, easy to handle and knot,<sup>38</sup> and, some would argue, biodegradable.<sup>39,40</sup> Most surgeons opt for the strongest material to which they have access. Comparative studies have shown that with all factors controlled for, using a standardized 4-0 locked cruciate repair, stainless steel was strongest (87.4 N), followed closely by FiberWire (Arthrex, Inc., Naples, Fla.) (80.5 N), then Ethibond (Ethicon, Inc., Somerville, N.J.) (65.6 N), then Prolene (Ethicon) (63.4 N), and finally Nylon (Ethicon) (46.7 N).<sup>31</sup> The size of suture is proportional to the strength of the repair, but a

**Table 1. Mechanism of Injury Chart**

Etiologic Factor	Model	Evidence	References
Trauma	Chicken flexor tendons	Crush injury, suture repair, sheath excision, and sublimis excision and immobilization cause violent adhesion formation	Lindsay and Thomson, 1960 <sup>21</sup>
	Rabbit flexor tendons	Combination of sheath and tendon injury with tendon suturing and immobilization	Matthews and Richards, 1976 <sup>22</sup>
Thermal energy	Chicken flexor tendons	Diathermy and carbon dioxide laser exposure to peritendinous tissue was directly proportional to adhesion formation	Hatano et al., 2000 <sup>23</sup>
Infection	Horse flexor tendons	Tenoscopic evidence of dense fibrous adhesions following tendon sheath infections	Bertone, 1995 <sup>24</sup>

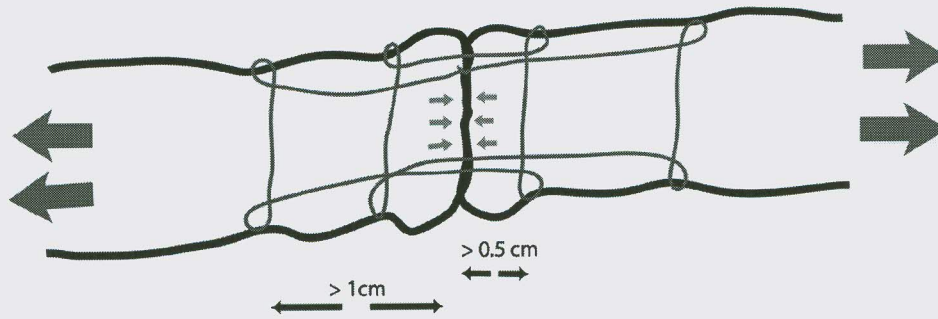


**Fig. 3.** Factors that influence tendon suture repair strength. Variables were deduced from ex vivo cadaver studies into (left) suture material with standardized 4-0 four-strand locked cruciate,<sup>31</sup> (second column) suture gauge with Ethibond as a standardized two-strand modified Kessler.<sup>32</sup> (Third column) Number of strands comparing Kessler (two-strand), cruciate (four-strand), Savage (six-strand),<sup>33</sup> and eight-strand Kessler<sup>34</sup> using standardized 4-0 Ethibond and 6-0 nylon epitendinous locking. (Fourth column) Suture configuration using standardized 4-0 FiberWire.<sup>35</sup> (Fifth column) Suture purchase using standardized 3-0 FiberWire locked cruciate and 6-0 interlocking horizontal mattress epitendinous suture.<sup>36</sup> (Right) Epitendinous suture using standardized 3-0 nylon and 5-0 nylon running epitendinous.<sup>37</sup>

reasonable size should be selected for the tendon size, to avoid overly damaging the tendon with needle size.<sup>32</sup> Multistrand repairs have gained popularity over double-strand repairs over the years, supported by a number of ex vivo and clinical studies (see Wu and Tang<sup>41</sup>). In average hands, the multistrand repair provides an extra 20 to 30 N of strength safeguard over a double-strand repair. The Savage six-strand repair<sup>42</sup> can give 81 percent excellent or good results in experienced hands but has failed to gain universal acceptance because of its complex nature.<sup>43</sup> The development and evolution of six-strand looped suture repairs by Wu and Tang has shown equivalent results with

simpler M and U configurations that benefit from fewer suture passes and balanced load across the cross-section of the tendon.<sup>41</sup>

Repair geometry is critically important to disperse load and offer a balanced repair. Performing an epitendinous suture in modified Kessler repairs has this stabilizing effect,<sup>37</sup> or the load dispersion effect can be produced by having the sutures placed asymmetrically.<sup>44</sup> For a given suture caliber, altering the configuration of a repair by asymmetrically staggering the strands of a four-strand Kessler repair offers a greater repair strength than a symmetrical four-strand Kessler repair.<sup>35</sup>

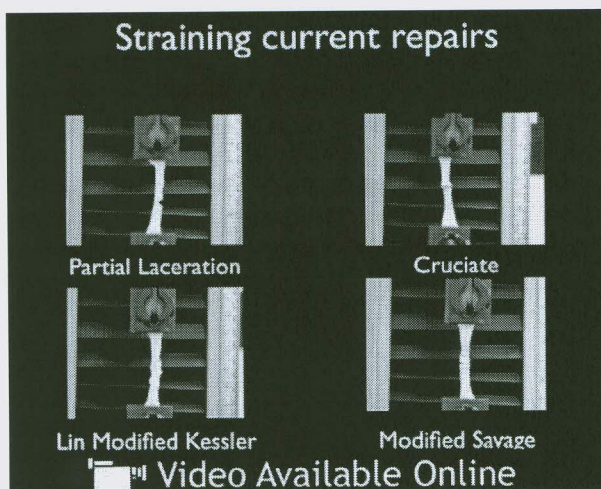


**Fig. 4.** Lin modified four-strand Kessler repair, one of the few dynamic repairs that offers compression on distraction through its continuous and asymmetric design. (See Lin GT. New suture techniques for flexor tendon repair. In: Saffar P, Amadio P, Foucher G, eds. *Current Practice in Hand Surgery*. London: Martin Dunitz Ltd; 1997:17–20.)

In our center, a four-strand repair favored and familiar to the individual operating surgeon is used, which is key to the repair being performed well and minimizing trauma from overzealous suture insertion.<sup>27</sup> We advocate as large a bite as possible of the tendon ends based on the findings of Kim et al.<sup>45</sup> My personal favored option is the Lin modified Kessler, which is a continuous asymmetric four-strand Kessler with a single continuous 3-0 Prolene suture that allows for distraction of the repair to result in compression of the repair

site<sup>46</sup> (Fig. 4). This dynamic repair will adjust its configuration according to the pull across the repair site; a characteristic that other repairs do not provide. (See Video, Supplemental Digital Content 3, which displays a straining of popular repairs. The repairs include a standardized four-strand cruciate repair, four-strand Savage repair, and four-strand Lin modified Kessler repair. All are pulled apart at the same rate and force using 4-0 Prolene. Partially divided tendon, 50 percent, is used as a comparison and repairs are pulled to failure. Note the gapping that occurs with cruciate and Savage compared with the Lin modified Kessler repair. Ultimately, partial tendon is stronger than the other displayed repair types, available in the “Related Videos” section of the full-text article on PRSJJournal.com or available at <http://links.lww.com/PRS/B159>.) In addition, I perform a simple 5-0 or 6-0 running epitendinous to stabilize the repair. The biomechanics of looped suture techniques<sup>36</sup> and barbed suture techniques<sup>48</sup> are also a current vogue but are not readily available to all hand centers.

It is worth highlighting that some studies and systematic reviews indicate the best functional outcomes have been achieved using two-strand repairs and early active mobilization or Kleinert type rehabilitation, with acceptably low rupture rates.<sup>49–52</sup> For progress to be made in tendon repair design, there has to be a move away from thinking about the number of cores, locking and grasping, to calculating the precise architecture of an optimal repair based on the tendon morphology, biology, and unique characteristics of each tendon injury.



**Video 3.** Supplemental Digital Content 3 displays a straining of popular repairs. The repairs include a standardized four-strand cruciate repair, four-strand Savage repair, and four-strand Lin modified Kessler repair. All are pulled apart at the same rate and force using 4-0 Prolene. Partially divided tendon, 50 percent, is used as a comparison and repairs are pulled to failure. Note the gapping that occurs with cruciate and Savage compared with the Lin modified Kessler repair. Ultimately, partial tendon is stronger than the other displayed repair types, available in the “Related Videos” section of the full-text article on PRSJJournal.com or available at <http://links.lww.com/PRS/B159>.

## TENDON EXPOSURE AND RETRIEVAL

Exposure for a tendon repair is usually down to operator preference. Some prefer Bruner’s zig-zag



**Video 4.** Supplemental Digital Content 4 displays a flexor tendon retrieval using minimal access technique, available in the “Related Videos” section of the full-text article on PRSJJournal.com or available at <http://links.lww.com/PRS/B160>.



**Video 5.** Supplemental Digital Content 5 displays a wide-awake flexor tendon operation, available in the “Related Videos” section of the full-text article on PRSJJournal.com or available at <http://links.lww.com/PRS/B161>.

approach,<sup>53</sup> and others prefer midlateral.<sup>54</sup> These exposures aim to minimize the possibility of scar across the joint crease. The midlateral approach minimizes scarring on the volar aspect of the digit and, when performed with distal extension, gives excellent exposure to the retracted distal ends of tendon. However, in neonates, its use should be guarded, as there have been reports that the lateral scar migrates volarly with growth and may cause flexion contractures.<sup>55</sup> To retrieve the proximal tendon end, if it is not readily visible, the natural creases can serve as vents to localize the divided tendon, and with the aid of a feeding catheter can be retracted back into the injury zone for repair.<sup>56</sup> (See Video, Supplemental Digital Content 4, which displays a flexor tendon retrieval using minimal access technique, available in the “Related Videos” section of the full-text article on PRSJJournal.com or available at <http://links.lww.com/PRS/B160>.) This has the benefit of minimizing surgical wounds and thus reduces the inflammatory response, adhesions, and scarring over the zone II region of the hand.<sup>16</sup>

### WIDE-AWAKE FLEXOR TENDON SURGERY

The convention of operating on tendons in a bloodless field is a privilege that most hand surgeons appreciate and requires an anesthetized patient or at the very least an upper arm block so that the patient can tolerate the tourniquet for the required duration of surgery. The benefits of this are a clean, bloodless operating field in which structures can be easily identified and

wounds can be extended. Therefore, operating on a patient without tourniquet is counterintuitive and goes against Bunnell’s philosophy.<sup>57</sup> To challenge dogma, Lalonde and others have published numerous series on one of the major paradigm shifts in hand surgery using wide-awake anesthesia.<sup>58–60</sup> More hand surgeons are turning to this option for simple hand surgery. The benefits are not immediately apparent, as the first few cases require one to become comfortable with minor amounts of blood in the operating field; however, with experience, the benefits and ease of this technique in the context of flexor tendon repairs are quite dramatic. (See Video, Supplemental Digital Content 5, which displays a wide-awake flexor tendon operation, available in the “Related Videos” section of the full-text article on PRSJJournal.com or available at <http://links.lww.com/PRS/B161>.) The ability to assess the repair intraoperatively for gapping, triggering, bowstringing, and smoothness of glide gives both the surgeon and patient confidence to proceed to an early active mobilization regimen.<sup>59</sup>

### PULLEY MANAGEMENT

The debate on whether to vent or not vent the pulleys depends largely on what degree of impingement can be seen by putting the digit through a full range of motion on the operating table. Despite the early studies by Doyle suggesting that the A2 and A4 pulley were sacrosanct,<sup>61</sup> Franko et al. showed that venting the A4 pulley, in the context of a repair close to the pulley, allowed an increase in excursion by 5 percent and



reduced the work of flexion compared with the nonvented A4 pulley.<sup>62</sup> The more detailed analysis by Tang and Xie has shown that 75 percent of the A2 and complete division of the A4 pulley can be performed without much detriment to the digital arc of motion.<sup>63</sup> In addition, clinical studies indicate that as many as 56 percent of zone II injuries would benefit from venting either the A2 or A4 pulley to allow for sufficient glide.<sup>64</sup>

### EVOLVING PRACTICE

The heterogeneity in the timing of the repairs, the demographics of the patient populations, mechanisms of injury, surgical approach, and surgical method make it difficult to define the factors that give the best outcomes. Incrementally evolving one's own practice, as Sirota-kova and Elliot have with flexor pollicis longus tendons, is one way of controlling for these confounding influences. The evolution of their flexor pollicis longus injury management has seen a stepwise improvement from using two-strand Kessler with simple peripheral sutures<sup>65</sup> to two-strand Kessler and Silfverskiöld peripheral sutures, to a double-stranded Kessler and Silfverskiöld peripheral suture,<sup>66</sup> to triple Tsuge repair with no peripheral suture,<sup>67</sup> and saw improvements from 70 to 75 to 75 percent good, excellent functional results and 17 to 8 to 0 percent and 0 percent rupture rates, respectively. Our own experience with repeated audit of our clinical activities in flexor tendons has seen incremental improvements from 1997 to 2013, with rupture rates dropping from 30 percent with two-strand Kessler repairs, epitendinous repair, and controlled active mobilization regimens, through to 17 percent with the introduction of hand therapist-led clinics, to rupture rates of 4 percent with the introduction of four-strand repairs.<sup>68</sup> Through the unit policy introduction of four-strand core repairs, circumferential epitendinous suture, along with consultant-led trauma lists, practitioner-led clinics, and dedicated tendon repair training days, our unit rupture rate was reduced to 0 percent in 2006. We also found that delaying the time of surgery beyond 7 days had a significant negative impact on patients' range of motion which, once ameliorated, led to better functional results. As such, there is great merit in performing in-house analysis and continual health care improvement in centers that manage flexor tendon injuries because of the heterogeneous patient demographics, surgical skill sets, and therapy provisions from unit to

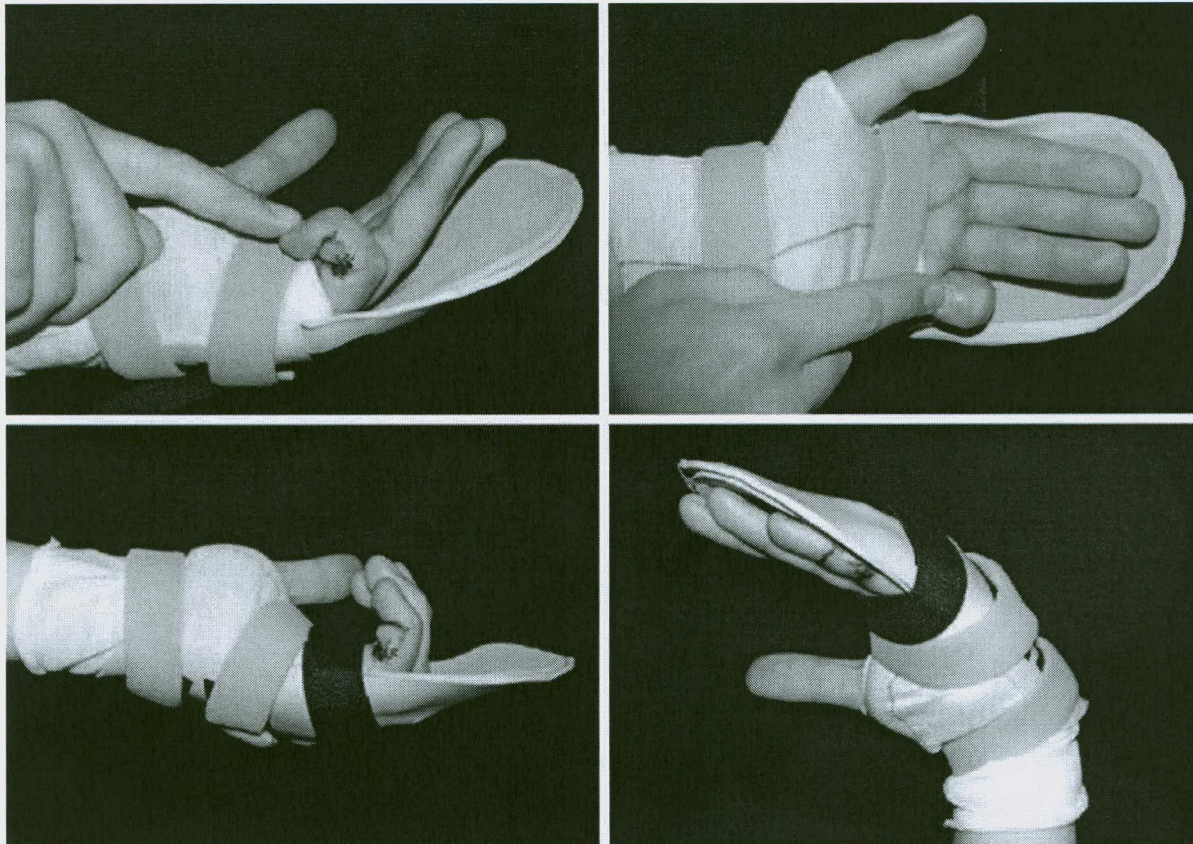
unit. Arguably, this is more relevant to individual centers than reported systematic reviews that pool results and give confounding outcomes.<sup>48-</sup>

### REHABILITATION

The evidence for the best rehabilitation regimen has been heavily debated over the years. Biologically, there is sound evidence that correlates well with clinical observations that tensile motion facilitates a reduction in inflammation,<sup>69</sup> up-regulation of collagen synthesis,<sup>70</sup> deposition of large-diameter fibrils,<sup>71</sup> and alignment of collagen fibrils.<sup>72</sup> As such, the argument for early active mobilization is scientifically quite compelling. Many reviews have now compared early active mobilization with passive mobilization and combined therapies, and the rupture rates are comparable but overall range of active movement is certainly better with the active regimens as indicated by level IIa evidence.<sup>73</sup> A review of electronic databases between 1970 and 2009 of 15 articles that reviewed rupture rates, range of motion, and quality of life identified that rupture rates were lowest in combined therapy regimens (controlled passive motion and passive flexion, active extension Kleinert and Duran) (2.3 percent) and highest in Kleinert-only (passive flexion and active extension) protocols (7.1 percent); however, finding the best functional results were obtained by early active mobilization or combined Kleinert and Duran protocols.<sup>74</sup> Studies into quality-of-life measures are still lacking.

There are a number of elements that in our experience appear to affect outcomes, including mechanism of injury, zone of injury, timing of surgery, caliber of repair, and age and characteristics of the patient, although this is not reflected in systematic review meta-analysis of complications following flexor tendon repair.<sup>6</sup> In each case, it is important that there is a degree of customization of therapy regimens to the needs of the patient, with close effective communication of the operative findings and surgery performed between the surgeon and therapists.

In Manchester, we have redesigned our rehabilitation regimens in line with increased repair strengths. Regimens must adhere to the principles of safety and promote maximum tendon glide. Active mobilization regimens for noncomplex repairs begin on the third to fifth days, in line with current thinking on the subsidence of postoperative edema.<sup>28</sup> To minimize the effects of edema on joint motion and to prevent tightening

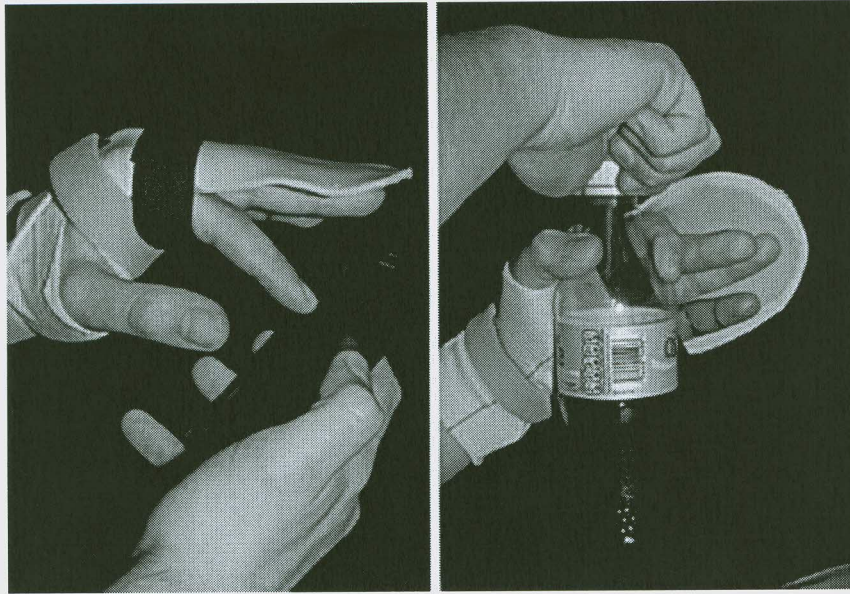


**Fig. 5.** Manchester short splint. (Above) Patients are taught to implement full passive range of flexion of the digit after 5 days, followed by (below, left) active flexion, performed with the wrist in 45 degrees' extension initiated at the distal interphalangeal joint in full fist. (Below, right) Full active extension of the interphalangeal joints is performed with the wrist in flexion to reduce force across the flexors.

of dorsal structures, the exercise sequence prioritizes full passive flexion stretching of the interphalangeal joints, maximizing passive digital flexion before initiating active motion. We have also radically evolved our splinting design for greater patient freedom. The Manchester short splint extends from the fingertips to the proximal wrist crease with a block to 30 degrees of metacarpophalangeal flexion. This position facilitates active motion initiated from the distal interphalangeal joint with the wrist in 45-degree extension to promote differential glide and minimize the work of flexion (Figs. 5 and 6). This splint also permits maximal wrist flexion, facilitating maximal active interphalangeal joint extension to reduce the risk of extension loss common in this type of injury. (See Video, Supplemental Digital Content 6, which displays a Manchester short splint and rehabilitation, available in the "Related Videos" section of the full-text article on PRSJournal.com or available at <http://links.lww.com/PRS/B162>.)

Early results are encouraging, with improvements in the arc of flexion and reduced interphalangeal joint extension loss.<sup>75</sup> Patients are required to adhere to a strict rehabilitation regimen, and compliance is an important factor in achieving good outcomes. Compliance differs according to age, understanding, mental health issues, and socioeconomic factors. Previous studies report high ruptures in those patients who use the hand for inappropriate functional activity.<sup>76</sup> Exclusion of the affected hand from function for 6 weeks is perhaps unrealistic in today's socioeconomic conditions, and we now instruct the patient in safe use of the hand with exclusion of the affected digit (Fig. 6, left).

A protective guard or boxing glove dressing may be used to add extra protection to the unpredictable activity of a child. For very young children, immobilization for 4 weeks in dressings does not appear to give rise to the same adhesion problems as for adults.<sup>77</sup>



**Fig. 6.** Patients are taught to exclude the injured digit but can use their hand for day-to-day tasks. This improves patient compliance with the splint.

### FUTURE DEVELOPMENTS

There are a number of technologies in development that may impact on the management of tendon injuries in the future. Biomolecules, gene therapies, and cell-based therapies have emerged from the biotechnology field and have been applied to tendon repairs. Transforming growth factor- $\beta$  and manipulation of these pathways have undergone preclinical study<sup>78</sup> but ultimately have not delivered in the clinical setting, possibly because of the heterogeneity of human populations posing too much of a challenge to



Video Available Online

**Video 6.** Supplemental Digital Content 6 displays a Manchester short splint and rehabilitation, available in the “Related Videos” section of the full-text article on PRSJournal.com or available at <http://links.lww.com/PRS/B162>.

single molecular therapies or single pathway manipulation approaches. Even multiple growth factor manipulation in solutions such as platelet-rich plasma have shown mixed results in augmenting flexor tendon repair. Some studies have shown enhancement at 2 weeks<sup>79</sup> and others have shown no significant improvement,<sup>80</sup> and the “silver bullet” approach to flexor tendon injuries is unlikely to be successful. As such, gene therapy and stem cell therapies are not likely to have a great impact on addressing issues arising from the standard simple divided tendon and repair. Adjuncts to improve glide may have a role, and carbodiimide-derived hyaluronic acid with or without lubricin<sup>81</sup> or using shear aggregated fibronectin tubes<sup>82</sup> certainly shows promise in animal studies, but cost-to-benefit consideration to clinical translation is likely to be a major hurdle. For therapies to replace damaged tendon, reseeding acellular tendon with stem cells<sup>83</sup> and using biomaterials as scaffolds<sup>84</sup> are popular research areas at present. In the future, use of individualized custom splints scanned to fit using three-dimensional printing may be commonplace in all hand therapy departments and has already been touted as a massive area of growth in the health care industry.<sup>85</sup>

### CONCLUSIONS

Minor gains in flexor tendon outcomes can be expected with the evolution of operating equipment, surgical devices, techniques, and

rehabilitation regimens, but until a major biological or technological disruptive technology arises that affects all forms of wound healing and repair processes, progress in this field will be modest. Until then, the pursuit of improving flexor tendon outcomes will continue to monopolize the research interests of many academic hand surgeons.

At present, using a combination of early surgical intervention, multistrand repairs, minimal access tendon surgery under wide-awake anesthesia, with the Manchester short splint regimen for rehabilitation, along with appropriate patient education is what we believe will improve the functional results in our patient population. This will have to be examined prospectively to provide better levels of supportive evidence; however, history dictates that evolution and innovation in this field are inevitable.

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#### PATIENT CONSENT

*The patient provided written consent for use of the patient's image.*

#### REFERENCES

1. Elliot D, Giesen T. Primary flexor tendon surgery: The search for a perfect result. *Hand Clin.* 2013;29:191–206.
2. RCS. Core skills in hand surgery. Available at: <http://www.rcseng.ac.uk/courses/course-search/hand-surgery.html>. Accessed January 9, 2014.
3. Lister GD, Kleinert HE, Kutz JE, Atasoy E. Primary flexor tendon repair followed by immediate controlled mobilization. *J Hand Surg Am.* 1977;2:441–451.
4. Sandow MJ, McMahon M. Active mobilisation following single cross grasp four-strand flexor tenorrhaphy (Adelaide repair). *J Hand Surg Eur Vol.* 2011;36:467–475.
5. Winter J. Hospital episode statistics: Main procedures and interventions. 2013. Available at: <http://www.hscic.gov.uk>. Accessed January 9, 2014.
6. Dy CJ, Hernandez-Soria A, Ma Y, Roberts TR, Daluiski A. Complications after flexor tendon repair: A systematic review and meta-analysis. *J Hand Surg Am.* 2012;37:543–551.e1.
7. Potenza AD. Tendon healing within the flexor digital sheath in the dog. *J Bone Joint Surg Am.* 1962;44:49–64.
8. Hanff G, Hagberg L. Prevention of restrictive adhesions with expanded polytetrafluoroethylene diffusible membrane following flexor tendon repair: An experimental study in rabbits. *J Hand Surg Am.* 1998;23:658–664.
9. Matthews P, Richards H. The repair reaction of flexor tendon within the digital sheath. *Hand* 1975;7:27–29.
10. Rank F, Eiken O, Bergenholtz A, Lundborg G, Erkel LJ. Flexor tendon specimens in organ cultures. *Scand J Plast Reconstr Surg.* 1980;14:179–183.
11. Manske PR, Lesker PA. Histologic evidence of intrinsic flexor tendon repair in various experimental animals: An in vitro study. *Clinic Orthop Relat Res.* 1984;182:297–304.
12. Manske PR, Lesker PA. Biochemical evidence of flexor tendon participation in the repair process: An in vitro study. *J Hand Surg Br.* 1984;9:117–120.
13. Lundborg G, Rank F, Heinau B. Intrinsic tendon healing: A new experimental model. *Scand J Plast Reconstr Surg.* 1985;19:113–117.
14. Kapacee Z, Richardson SH, Lu Y, et al. Tension is required for fibroblast formation. *Matrix Biol.* 2008;27:371–375.
15. Evans CE, Trail IA. Fibroblast-like cells from tendons differ from skin fibroblasts in their ability to form three-dimensional structures in vitro. *J Hand Surg Br.* 1998;23:633–641.
16. Wong JK, Lui YH, Kapacee Z, Kadler KE, Ferguson MW, McGrouther DA. The cellular biology of flexor tendon adhesion formation: An old problem in a new paradigm. *Am J Pathol.* 2009;175:1938–1951.
17. Chen CH, Cao Y, Wu YF, Bais AJ, Gao JS, Tang JB. Tendon healing in vivo: Gene expression and production of multiple growth factors in early tendon healing period. *J Hand Surg Am.* 2008;33:1834–1842.
18. Guimberteau JC, Delage JP, McGrouther DA, Wong JK. The microvacuolar system: How connective tissue sliding works. *J Hand Surg Eur Vol.* 2010;35:614–622.
19. Lindsay WK, McDougall EP. Direct digital flexor tendon repair. *Plast Reconstr Surg Transplant Bull.* 1960;26:613–621.
20. Taylor SH, Al-Youha S, Van Agtmael T, et al. Tendon is covered by a basement membrane epithelium that is required for cell retention and the prevention of adhesion formation. *PLoS One* 2011;6:e16337.
21. Lindsay WK, Thomson HG. Digital flexor tendons: An experimental study. Part I. The significance of each component of the flexor mechanism in tendon healing. *Br J Plast Surg.* 1960;12:289–316.
22. Matthews P, Richards H. Factors in the adherence of flexor tendon after repair: An experimental study in the rabbit. *J Bone Joint Surg Br.* 1976;58:230–236.
23. Hatano I, Suga T, Diao E, Peimer CA, Howard C. Adhesions from flexor tendon surgery: An animal study comparing surgical techniques. *J Hand Surg Am.* 2000;25:252–259.
24. Bertone AL. Tendon lacerations. *Vet Clin North Am Equine Pract.* 1995;11:293–314.
25. Starnes T, Saunders RJ, Means KR Jr. Clinical outcomes of zone II flexor tendon repair depending on mechanism of injury. *J Hand Surg Am.* 2012;37:2532–2540.
26. Zhao C, Amadio PC, Momose T, Couvreur P, Zobitz ME, An KN. The effect of suture technique on adhesion formation after flexor tendon repair for partial lacerations in a canine model. *J Trauma* 2001;51:917–921.
27. Wong JK, Alyouha S, Kadler KE, Ferguson MW, McGrouther DA. The cell biology of suturing tendons. *Matrix Biol.* 2010;29:525–536.
28. Wu YF, Tang JB. Tendon healing, edema, and resistance to flexor tendon gliding: Clinical implications. *Hand Clin.* 2013;29:167–178.

29. Small JO, Brennen MD, Colville J. Early active mobilisation following flexor tendon repair in zone 2. *J Hand Surg Br*. 1989;14:383–391.
30. Strickland JW. Flexor tendon injuries: I. Foundations of treatment. *J Am Acad Orthop Surg*. 1995;3:44–54.
31. Lawrence TM, Davis TR. A biomechanical analysis of suture materials and their influence on a four-strand flexor tendon repair. *J Hand Surg Am*. 2005;30:836–841.
32. Taras JS, Raphael JS, Marczyk SC, Bauerle WB. Evaluation of suture caliber in flexor tendon repair. *J Hand Surg Am*. 2001;26:1100–1104.
33. Barrie KA, Wolfe SW, Shean C, Shenbagamurthi D, Slade JF III, Panjabi MM. A biomechanical comparison of multi-strand flexor tendon repairs using an in situ testing model. *J Hand Surg Am*. 2000;25:499–506.
34. Winters SC, Seiler JG III, Woo SL, Gelberman RH. Suture methods for flexor tendon repair: A biomechanical analysis during the first six weeks following repair. *Ann Chirurgie Main Memb Super*. 1997;16:229–234.
35. Haimovici L, Papafragkou S, Lee W, Dagum A, Hurst LC. The impact of fiberwire, fiberloop, and locking suture configuration on flexor tendon repairs. *Ann Plast Surg*. 2012;69:468–470.
36. Lee SK, Goldstein RY, Zingman A, Terranova C, Nasser P, Hausman MR. The effects of core suture purchase on the biomechanical characteristics of a multistrand locking flexor tendon repair: A cadaveric study. *J Hand Surg Am*. 2010;35:1165–1171.
37. de Wit T, Walbeehm ET, Hovius SE, McGrouther DA. The mechanical interaction between three geometric types of nylon core suture and a running epitendon suture in repair of porcine flexor tendons. *J Hand Surg Eur Vol*. 2013;38:788–794.
38. Trail IA, Powell ES, Noble J. An evaluation of suture materials used in tendon surgery. *J Hand Surg Br*. 1989;14:422–427.
39. Caulfield RH, Maleki-Tabrizi A, Patel H, Coldham F, Mee S, Nanchahal J. Comparison of zones 1 to 4 flexor tendon repairs using absorbable and unabsorbable four-strand core sutures. *J Hand Surg Eur Vol*. 2008;33:412–417.
40. Kang HJ, Lee DC, Kim JS, Ki SH, Roh SY, Yang JW. Flexor tenorrhaphy using absorbable suture materials. *Arch Plast Surg*. 2012;39:397–403.
41. Wu YF, Tang JB. Recent developments in flexor tendon repair techniques and factors influencing strength of the tendon repair. *J Hand Surg Eur Vol*. 2014;39:6–19.
42. Savage R. In vitro studies of a new method of flexor tendon repair. *J Hand Surg Br*. 1985;10:135–141.
43. Savage R, Risitano G. Flexor tendon repair using a “six strand” method of repair and early active mobilisation. *J Hand Surg Br*. 1989;14:396–399.
44. Wu YF, Tang JB. The effect of asymmetric core suture purchase on gap resistance of tendon repair in linear cyclic loading. *J Hand Surg Am*. 2014;39:910–918.
45. Kim JB, de Wit T, Hovius SE, McGrouther DA, Walbeehm ET. What is the significance of tendon suture purchase? *J Hand Surg Eur Vol*. 2009;34:497–502.
46. Lin GT. New suture techniques for flexor tendon repair. In: Saffar P, Amadio P, Foucher G, eds. *Current Practice in Hand Surgery*. London: Martin Dunitz Ltd; 1997:17–20.
47. Tang JB, Shi D, Gu YQ, Chen JC, Zhou B. Double and multiple looped suture tendon repair. *J Hand Surg Br*. 1994;19:699–703.
48. Peltz TS, Haddad R, Scougall PJ, Gianoutsos MP, Bertollo N, Walsh WR. Performance of a knotless four-strand flexor tendon repair with a unidirectional barbed suture device: A dynamic ex vivo comparison. *J Hand Surg Eur Vol*. 2014;39:30–39.
49. Bainbridge LC, Robertson C, Gillies D, Elliot D. A comparison of post-operative mobilization of flexor tendon repairs with “passive flexion-active extension” and “controlled active motion” techniques. *J Hand Surg Br*. 1994;19:517–521.
50. Kitis PT, Buker N, Kara IG. Comparison of two methods of controlled mobilisation of repaired flexor tendons in zone 2. *Scand J Plast Reconstr Surg Hand Surg*. 2009;43:160–165.
51. Saldana MJ, Chow JA, Gerbino P II, Westerbeck P, Schacherer TG. Further experience in rehabilitation of zone II flexor tendon repair with dynamic traction splinting. *Plast Reconstr Surg*. 1991;87:543–546.
52. Hardwicke JT, Tan JJ, Foster MA, Titley OG. A systematic review of 2-strand versus multistrand core suture techniques and functional outcome after digital flexor tendon repair. *J Hand Surg Am*. 2014;39:686–695.e2.
53. Bruner JM. The zig-zag volar-digital incision for flexor-tendon surgery. *Plast Reconstr Surg*. 1967;40:571–574.
54. Hall RF Jr, Vliegenthart DH. A modified midlateral incision for volar approach to the digit. *J Hand Surg Br*. 1986;11:195–197.
55. Kavouksorian CA, Noone RB. Flexor tendon repair in the neonate. *Ann Plast Surg*. 1982;9:415–418.
56. Wong J, McGrouther DA. Minimizing trauma over ‘no man’s land’ with flexor tendon retrieval. *J Hand Surg Eur Vol*. 2014;39:1004–1006.
57. Bunnell S. *Surgery of the Hand*. 2nd ed. Philadelphia: Lippincott; 1948.
58. Lalonde DH. Wide-awake flexor tendon repair. *Plast Reconstr Surg*. 2009;123:623–625.
59. Higgins A, Lalonde DH, Bell M, McKee D, Lalonde JF. Avoiding flexor tendon repair rupture with intraoperative total active movement examination. *Plast Reconstr Surg*. 2010;126:941–945.
60. Lalonde DH, Martin AL. Wide-awake flexor tendon repair and early tendon mobilization in zones 1 and 2. *Hand Clin*. 2013;29:207–213.
61. Doyle JR. Anatomy of the flexor tendon sheath and pulley system: A current review. *J Hand Surg Am*. 1989;14:349–351.
62. Franko OI, Lee NM, Finneran JJ, et al. Quantification of partial or complete A4 pulley release with FDP repair in cadaveric tendons. *J Hand Surg Am*. 2011;36:439–445.
63. Tang JB, Xie RG. Effect of A3 pulley and adjacent sheath integrity on tendon excursion and bowstringing. *J Hand Surg Am*. 2001;26:855–861.
64. Kwai Ben I, Elliot D. “Venting” or partial lateral release of the A2 and A4 pulleys after repair of zone 2 flexor tendon injuries. *J Hand Surg Br*. 1998;23:649–654.
65. Sirotakova M, Elliot D. Early active mobilization of primary repairs of the flexor pollicis longus tendon. *J Hand Surg Br*. 1999;24:647–653.
66. Sirotakova M, Elliot D. Early active mobilization of primary repairs of the flexor pollicis longus tendon with two Kessler two-strand core sutures and a strengthened circumferential suture. *J Hand Surg Br*. 2004;29:531–535.
67. Giesen T, Sirotakova M, Copsey AJ, Elliot D. Flexor pollicis longus primary repair: Further experience with the Tang technique and controlled active mobilization. *J Hand Surg Eur Vol*. 2009;34:758–761.
68. Wong J, Peck F, Highton L, et al. The evolving practice of flexor tendon management in a tertiary hand centre. Paper presented at: British Society for Surgery of the Hand Scientific Meeting, April 21, 2010; Manchester, United Kingdom.

69. Iwuagwu FC, McGrouther DA. Early cellular response in tendon injury: The effect of loading. *Plast Reconstr Surg.* 1998;102:2064–2071.
70. Li F, Li B, Wang QM, Wang JH. Cell shape regulates collagen type I expression in human tendon fibroblasts. *Cell Motil Cytoskeleton* 2008;65:332–341.
71. Matthew C, Moore MJ, Campbell L. A quantitative ultra-structural study of collagen fibril formation in the healing extensor digitorum longus tendon of the rat. *J Hand Surg Br.* 1987;12:313–320.
72. Connizzo BK, Yannascoli SM, Soslowsky LJ. Structure-function relationships of postnatal tendon development: A parallel to healing. *Matrix Biol.* 2013;32:106–116.
73. Starr HM, Snoddy M, Hammond KE, Seiler JG III. Flexor tendon repair rehabilitation protocols: A systematic review. *J Hand Surg Am.* 2013;38:1712-7.e1–14.
74. Chesney A, Chauhan A, Kattan A, Farrokhyar F, Thoma A. Systematic review of flexor tendon rehabilitation protocols in zone II of the hand. *Plast Reconstr Surg.* 2011;127:1583–1592.
75. Peck F. The rehabilitation of flexor tendon injuries in zone 2. *ifssh ezine* 2014;4:32–37.
76. Harris SB, Harris D, Foster AJ, Elliot D. The aetiology of acute rupture of flexor tendon repairs in zones 1 and 2 of the fingers during early mobilization. *J Hand Surg Br.* 1999;24:275–280.
77. Nietosvaara Y, Lindfors NC, Palmu S, Rautakorpi S, Ristaniemi N. Flexor tendon injuries in pediatric patients. *J Hand Surg Am.* 2007;32:1549–1557.
78. Bates SJ, Morrow E, Zhang AY, Pham H, Longaker MT, Chang J. Mannose-6-phosphate, an inhibitor of transforming growth factor-beta, improves range of motion after flexor tendon repair. *J Bone Joint Surg Am.* 2006;88:2465–2472.
79. Sato D, Takahara M, Narita A, et al. Effect of platelet-rich plasma with fibrin matrix on healing of intrasynovial flexor tendons. *J Hand Surg Am.* 2012;37:1356–1363.
80. Kollitz KM, Parsons EM, Weaver MS, Huang JI. Platelet-rich plasma for zone II flexor tendon repair. *Hand (NY)* 2014;9:217–224.
81. Zhao C, Sun YL, Kirk RL, et al. Effects of a lubricin-containing compound on the results of flexor tendon repair in a canine model in vivo. *J Bone Joint Surg Am.* 2010;92:1453–1461.
82. Branford OA, Mudera V, Brown RA, McGrouther DA, Grobbelaar AO. A novel biomimetic material for engineering postsurgical adhesion using the injured digital flexor tendon-synovial complex as an in vivo model. *Plast Reconstr Surg.* 2008;121:781–793.
83. Thorfinn J, Angelidis IK, Gigliello L, Pham HM, Lindsey D, Chang J. Bioreactor optimization of tissue engineered rabbit flexor tendons in vivo. *J Hand Surg Eur Vol.* 2012;37:109–114.
84. Bosworth LA, Alam N, Wong JK, Downes S. Investigation of 2D and 3D electrospun scaffolds intended for tendon repair. *J Mater Sci Mater Med.* 2013;24:1605–1614.
85. Paterson AM, Bibb RJ, Campbell RI. Evaluation of a digitised splinting approach with multiple-material functionality using additive manufacturing technologies. Paper presented at: Twenty-Third Annual International Solid Freeform Fabrication Symposium; August 6–11, 2011; Austin, Texas.